

## Role of Bcl-x<sub>L</sub> in Ionizing Radiation-Induced Invasion of Cancer Cells

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### 1. Introduction

Although radiation therapy is a major therapeutic modality for cancer treatment, previous reports have suggested that ionizing radiation (IR) can promote the invasive and metastatic potential of cancer cells [1]. Successful invasion of cancer cells requires their resistance to apoptosis once the cell has detached from the primary tumor tissue. Bcl-2 family of genes plays a key role in the regulation of apoptotic cell death [2]. Recent studies have revealed that overexpression of Bcl-2, Bcl-w, and Bcl-x<sub>L</sub> were associated with tumor progression [2-4]. This study investigated whether Bcl-x<sub>L</sub> can mediate the IR-induced invasion of lung cancer cells.

### 2. Methods and Results

#### 2.1 Cell culture

A549 human lung adenocarcinoma cells were obtained from American Type Culture Collection and grown in RPMI 1640 supplemented with 10% heat-inactivated fetal bovine serum and gentamicin (50 µg/ml) at 37°C in a humidified atmosphere containing 5% CO<sub>2</sub>.

#### 2.2 Irradiation

Cells were plated in 60-mm dishes and incubated at 37°C under humidified 5% CO<sub>2</sub> and 95% air in culture medium until 70~80% confluent. Cells were then exposed to γ-rays from a <sup>137</sup>Cs γ-ray source [Atomic Energy of Canada] at a dose rate of 3.06 Gy/min.

#### 2.3 Invasion and migration assays

Invasion of A549 cell was measured by the invasion of cells through Matrigel-coated transwell inserts (Corning Inc., Corning, NY). Briefly, transwell inserts with 8-µm pore size were coated with Matrigel (1 mg/ml). Cells were trypsinized, and 200 µl of cell suspension (2×10<sup>5</sup> cells/ml) were added to the Matrigel-coated chamber; the lower compartment was filled with RPMI 1640 conditioned media (1 ml). After incubation for 16~20 h at 37°C, cells on the surface of Matrigel-coated polycarbonate membrane (non-invading cells) were removed with a cotton swab and the migrating cells remaining on the bottom part of the filters were fixed and stained with Diff-Quick kit (Fisher Scientific, Pittsburgh, PA) and then counted under a microscope.

Migration assays were done using the same procedure but with uncoated polycarbonate filters.

#### 2.4 Western blot

Cells were lysed in ice-cold lysis buffer containing 1% Triton X-100 in 20 mM Tris-HCl (pH 7.5) and protease inhibitor cocktail (Sigma-Aldrich, St. Louis, MO). Lysates were then centrifuged at 14,000 rpm for 20 minutes, equal amounts of proteins (30~50 µg) were separated by 12% SDS-PAGE. The proteins were then electrotransferred to immobilon membranes (Millipore, Bedford, MA), which were subsequently blotted using the indicated antibodies and visualized by chemiluminescence (ECL; Amersham, Arlington Heights, IL).

### 3. Results

#### 3.1 IR and Bcl-x<sub>L</sub> promote the invasiveness of lung cancer cells

IR increased invasive potential of lung cancer cells in a dose-dependent manner and the highest invasiveness observed at 10 Gy of γ-rays (Fig. 1A). Bcl-x<sub>L</sub> expression was increased in time dependent manner by exposure of IR (Fig. 1B). To determine whether Bcl-x<sub>L</sub> influences the invasiveness of lung cancer cells, Bcl-x<sub>L</sub> was overexpressed in A549 cells. As expected, overexpression of Bcl-x<sub>L</sub> also promoted their invasion but did not show synergy effect (Fig. 1C). Taken together, IR and Bcl-x<sub>L</sub> appears to promote the invasion of A549 cells by same mechanism.

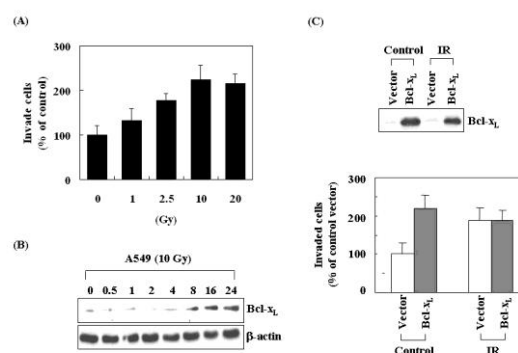


Fig. 1. IR and Bcl-x<sub>L</sub> promote the invasive potential of A549 cells. A549 cells were irradiated with 10 Gy of γ-rays. At the end of 24 hr of incubation, Invasion assays (A), and Western blot (B) were performed. Bcl-x<sub>L</sub> overexpression was introduced into A549, and then irradiated with or without 10 Gy of γ-rays. Invasion assays (C) were performed.

### 3.2 MAPK is involved in IR-induced invasion

To investigate whether MAPK acts in IR-induced signaling pathways, we performed Western blot. As indicated in Fig. 2A, IR significantly enhanced the phosphorylation of Akt and p38 and MMP-2 secretion. To further confirm the signaling molecules involved in the IR-induced responses, the cells were irradiated with 10 Gy of  $\gamma$ -rays in the presence or absence of LY294002, SB202190, PD98059, and SP600125 which inhibit PI3K, p38 MAPK, ERK, and JNK, respectively. IR-induced cell invasion was effectively attenuated by the addition of LY294002 (Fig. 2B).

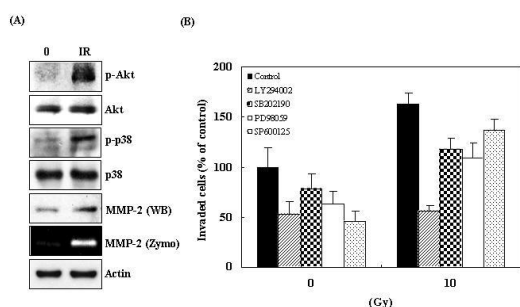


Fig. 2. MAPK is involved in IR-induced invasion. A549 cells were irradiated with 10 Gy of  $\gamma$ -rays. At the end of 24 hr of incubation, Western blot was performed (A), A549 Cells were pretreated with PI3K inhibitor (LY294002, 5  $\mu$ M), p38 inhibitor (SB202190, 5  $\mu$ M), Erk inhibitor (PD98059, 5  $\mu$ M), JNK inhibitor (SP600125, 5  $\mu$ M) for 30 min, and then irradiated with 10 Gy of  $\gamma$ -rays. At the end of 24 hr of incubation, Invasion assays were performed (B).

### 3.3 MAPK is involved in Bcl-x<sub>L</sub> induced invasion

As shown in Fig. 3A, Bcl-x<sub>L</sub> overexpression increased phosphorylation of Akt and p38 and MMP-2 secretion. Moreover, Bcl-x<sub>L</sub> failed to increase cell invasion when A549 cells were treated with LY294002 (Fig. 3B). These results suggest that Bcl-x<sub>L</sub> may mediate IR-induced invasion of A549 through the up-regulated PI3K/Akt pathway.

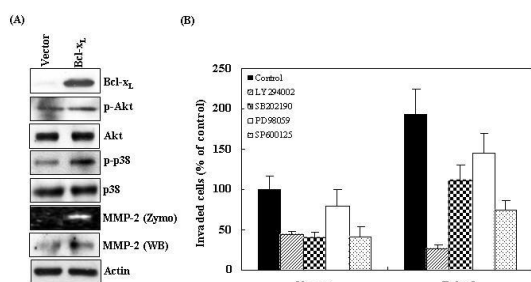


Fig. 3. MAPK is involved in Bcl-x<sub>L</sub>-induced invasion. (A) A549 cells were irradiated with 10 Gy of  $\gamma$ -rays. At the end of

24 hr of incubation, Western blot was performed (A), Cells were pretreated with PI3K inhibitor (LY294002, 5  $\mu$ M), p38 inhibitor (SB202190, 5  $\mu$ M), Erk inhibitor (PD98059, 5  $\mu$ M), JNK inhibitor (SP600125, 5  $\mu$ M) for 30 min, and then irradiated with 10 Gy of  $\gamma$ -rays. At the end of 24 hr of incubation, Invasion assays were performed (B).

### 3.4 Bcl-x<sub>L</sub> is necessary for IR-induced invasion

To determine whether Bcl-x<sub>L</sub> expression was crucial to the invasion, the levels of Bcl-x<sub>L</sub> were reduced by RNA interference. As shown in Fig. 4A, reduced Bcl-x<sub>L</sub> expression inhibited the IR-induced invasion compared with nonsilencing control cells. Moreover, IR-induced phosphorylation of Akt and p38 and MMP-2 secretion were reduced when the cells were treated with Bcl-x<sub>L</sub> siRNA (Fig. 4B).

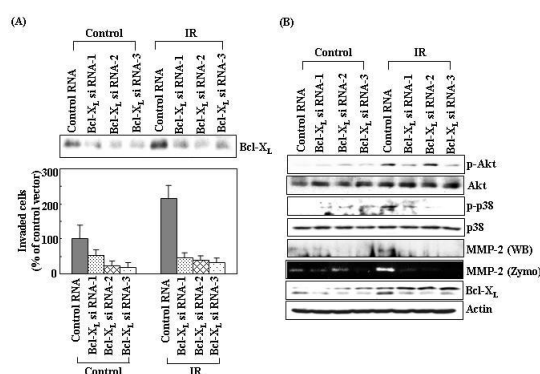


Fig. 4. Bcl-x<sub>L</sub> mediates IR-induced invasion. Bcl-x<sub>L</sub> siRNA was introduced into A549, and then irradiated with 10 Gy of  $\gamma$ -rays. After 24 hr of incubation, Invasion assay (A) and Western blot (B) were performed.

## 3. Conclusions

In the present study, we showed that Bcl-x<sub>L</sub> is a critical mediator of IR-induced invasion through the up-regulated phosphorylation of Akt and p38 pathway and MMP-2 secretion.

## REFERENCES

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